

Thyroid Follicular Cell Neoplasms in Multistep Carcinogenesis

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Abstract

The present review summarizes current knowledge on the alterations of molecular genetics and epigenetics of sporadic thyroid follicular cell neoplasms and the relationship of the alterations with multistep carcinogenesis theory. Approximately 50% of follicular thyroid carcinomas have mutations in RAS family genes or PAX8/PPARy gene rearrangements. These mutations are found in a mutually exclusive manner, which suggests that follicular thyroid carcinomas develop via two different initiating mechanisms. BRAF gene mutations and RET/PTC and NTRK1 rearrangements that involve the mitogen-activated protein kinase pathway are found in the majority of papillary thyroid carcinoma. These mutations are also found in a mutually exclusive manner and contribute to the initiation of the transformation from normal follicular cells to papillary thyroid carcinoma. TP53 mutations are almost exclusively found in poorly differentiated carcinoma and undifferentiated carcinoma and play an essential role in tumor progression. Aberrant methylation of oncogenes and suppressor oncogenes such as the PTEN gene also causes tumor progression. Growth factors (FGF, FGFR, MET, EGFR and VGEF), cell cycle regulators (cyclin D1, RB, p16, p21 and p27) and adhesion molecules (E-cadherin and fibronectin) activate the mitogenic signaling pathways and have impacts on the initiation, promotion and progression of thyroid carcinomas.

Keywords: Thyroid, follicular cells, carcinoma, gene, multistep carcinogenesis

Introduction

Two theories have been proposed regarding the thyroid follicular cell carcinogenesis: the fetal cell carcinogenesis theory proposed by Takano T and the more common multistep carcinogenesis theory by Vogelstein B et al. (1-6). As predisposing conditions for thyroid follicular cell carcinomas, various risk factors including nutritional (iodine intake), environmental (radiation exposure) and genetic (inherited tumor syndromes and familial non-medullary thyroid carcinoma) backgrounds have been reported. According to the multistep carcinogenesis theory, carcinogenesis is a sequential process involving alterations of

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multiple genes and epigenetics (4-6). The number of genes and the number of corresponding lesions have not been fully elucidated in thyroid follicular cell carcinogenesis, while as many as 10 or more sequential alterations have been shown to occur in colorectal carcinoma, in accordance with the multistep carcinogenesis theory (7-16). Analysis of thyroid carcinomas has disclosed accumulation of the changes of growth factors, cell cycle regulators and adhesion molecules, which activate mitogenic signaling pathways, in addition to genetic and epigenetic alterations of multiple genes. The present review summarizes current knowledge on the alterations of molecular genetics and epigenetics of sporadic (nonhereditary) thyroid follicular cell neoplasms.

Table 1: Hereditary (familial) thyroid carcinomas

C Cell Origin

- A) Syndromic types
 - Multiple endocrine neoplasia type 2 (MEN 2a/2b) (*RET* gene)
- B) Non-syndromic types

Familial Medullary Thyroid Carcinoma (RET gene)

Follicular Cell Origin

- A) Syndromic types
 - 1. Familial adenomatous polyposis (FAP) (APC gene)
 - 2. Cowden syndrome (PTEN hamartoma tumor syndrome) (*PTEN* gene)
 - 3. Multiple endocrine neoplasia type 1 (MEN 1) (*MEN1* gene)
 - 4. Carney complex (PRKAR1A gene)
 - 5. Werner syndrome (*WRN* gene)
 - 6. Li-Fraumeni syndrome (*TP53* gene)
- B) Non-syndromic types
 - Familial non-medullary thyroid carcinoma
 - 1. Familial papillary thyroid carcinoma (PTC) with papillary renal cell neoplasia (1q21)
 - 2. Familial PTC with oxyphilia (19p13.2)
 - 3. Familial PTC without oxyphilia (19p13.2)
 - 4. Familial multinodular goiter with PTC (14q)
 - 5. Familial PTC (2q21)

1. Hereditary Thyroid Carcinomas

Hereditary (familial) thyroid carcinoma can arise from both follicular cells and C cells of thyroid (Table 1). The majority of

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Genetic changes/ Histo-types	<i>RET/PTC</i> rearrangements	<i>BRAF</i> (V600E) mutations	<i>RAS</i> mutations	<i>PAX8/PPARγ</i> rearrangements	<i>CTNNB1</i> (β catenin) mutation	TP53 mutations	Ki-67 Labeling Index
FA	0%	0%	17-43%	8%	0%	0%	<3%
PTC	13-43%	30-69%	0-21%	0.3%	0%	0-5%	${<}10\%, usually {<}5\%$
FTC	0%	0%	40-53%	39%	0%	0-9%	<10%, usually <5%
PDC	0-13%	0-13%	18-55%	0%	25%	17-38%	10-30%
UC	0%	10-35%	4-60%	0%	66%	60-88%	>30%, usually >50%

Table 2: Genetic alterations in thyroid neoplasms and Ki-67 proliferation index*

FA: Follicular adenoma; PTC: Papillary thyroid carcinoma; FTC: Follicular thyroid carcinoma; PDC: Poorly differentiated carcinoma;

UC: Undifferentiated carcinoma; *modified from Kondo T et al., Nikiforov Y et al., Soares P et al., Omur O et al., Freitas BC et al. and Kakudo K et al.

thyroid carcinomas of follicular cell origin are non-hereditary (sporadic), however a 3% to 10% risk of well-differentiated thyroid carcinoma (WDC) has been documented in first-degree relatives of patients with WDC (17-22). WDC can be a minor component of several known hereditary tumor syndromes, such as Cowden syndrome (PTEN gene), familial adenomatous polyposis (APC gene), Carney complex (PRKAR1a gene) and Werner syndrome (WRN gene) (7, 13, 19, 21, 22). Inactivation of the PTEN gene through methylation has recently been found to be a common molecular event in follicular neoplasms (23-27). However, these responsible gene mutations are rare in sporadic PTC and follicular thyroid carcinoma (FTC) (28). In addition to those hereditary cancer syndromes, several susceptible gene loci have been identified in familial non-medullary thyroid carcinomas of non-syndromic type. Chromosomal locations identified in patients with familial thyroid carcinomas are 1q21, 2q21, 14q, and 19p13.2 (Table 1) (19, 21). However, majority of the sporadic thyroid carcinomas do not harbor allelic losses and DNA copy number changes at these loci (29-35).

2. Genetic Alterations in Sporadic Thyroid Neoplasms

PTC and FTC are WDC of follicular cell origin and constitute more than 90% of all types of thyroid malignancy. Although they are derived from the same follicular cells, they have different morphological features, biological behaviors and genetic alterations (7-14, 21, 33, 36). Four different gene mutations have been identified in PTC and FTC, including BRAF and RAS point mutations, and RET/PTC and PAX8/PPARy rearrangements. Poorly differentiated carcinoma (PDC) and undifferentiated carcinoma (UC) may occur either de novo or as a progression from WDC, and the minor components of PTC or FTC in PDC and UC are accepted as evidence of progression from WDC to PDC or UC (7, 11, 12, 16, 21, 37). Overexpression of cyclin D1, decreased expression of p27, and inactivation of PTEN, CTNNB1 (β-catenin) and TP53 genes have been reported to be responsible for the progression from WDC to PDC or UC (8, 11, 16, 24, 37-45) (Fig. 1).

a. Molecular Genetics of Follicular Neoplasms (Follicular Tumors)

Follicular neoplasms are a group of thyroid tumors that display a follicular growth pattern, and include follicular adenoma (FA), FTC and follicular variant of PTC. Gain-of-function mutations of *RAS* genes (*NRAS*, *KRAS* and *HRAS*) have been reported in all types of follicular neoplasm at various frequencies (46-55). Their prevalence is higher in FTC than in PTC, and in FTC than in FA (Table 2) (46-55). Some investigators concluded that *RAS* mutations may be important genetic alterations in the

initiation of thyroid follicular cell carcinogenesis because these mutations are present in all types of follicular neoplasm including FA, FTC, PDC and UC (47), while others concluded that N-RAS codon 61 mutation in FTC causes tumor progression and could be an important predictive factor for distant metastasis and a higher risk of anaplastic transformation (49, 50, 52, 54, 55). The peroxisome proliferator-activated receptor-gamma ($PPAR\gamma$) is a transcription factor essential for thyroid gland development, and the paired-box gene 8 (PAX8) is a member of the steroid/thyroid nuclear receptor family, and their fusion gene (PAX8/PPARy rearrangement) has been found in FA and FTC, but not in PTC, PDC and UC (9, 56-58) (Table 2). The two most common mutations in FTCs, PAX8/PPARy rearrangements and RAS point mutations, are seen in a mutually exclusive manner, which indicates that FTC could be initiated by two different mechanisms: either RAS point mutations or PAX8/PPARy rearrangements (9, 51).

b. Molecular Genetics of Papillary Thyroid Carcinoma

PTC is the most common histological type of thyroid malignancy globally and accounts for approximately 88% of all thyroid carcinomas in Japan (21, 36, 59). The most common genetic alterations such as in BRAF and RAS genes found in PTC are gain-of-function mutations that activate the mitogen-activated protein kinase (MAPK) pathway, which is responsible for thyroid tumorigenesis. The hotspot mutation, BRAF(V600E), is frequently detected in PTC (36-69%), in contrast to its absence in follicular neoplasms (benign FAs and malignant FTCs) (60-66) (Table 2). BRAF(V600E) mutation has been reported to be highly prevalent in PTC with a papillary and mixed papillary and follicular growth pattern, and BRAF(K601E) mutation is restricted to the follicular variant of PTC, suggesting possible genotype-phenotype correlations (62, 67). High frequency of BRAF(V600E) mutation has been reported in PTCs from early to late advanced stages and Ugolini et al. reported that BRAF(V600E) mutation was found in 17.6% of incidentally identified micro-PTCs and 38.3% of clinically identified micro-PTCs, indicating that this mutation is an early event in PTC tumorigenesis (14, 68-71). While BRAF mutations are found in up to 13% of PDC and 35% of UC, it is concluded that BRAF mutation-positive PTC may be more prone to PDC or UC transformation than PTC without BRAF mutation (66, 67-74). As BRAF mutation is highly prevalent (95%) in metastatic tumors from radioactive iodine-refractory PTC, these studies concluded that this mutation has prognostic implications (69, 71-75). However, conflicting data have been reported by some other investigators and BRAF(V600E) mutation has been suggested as a secondary subclonal change rather than a primary event in thyroid carcinogenesis (76-78).

The RET proto-oncogene is located on chromosome 10q11.2 and encodes a cell membrane receptor tyrosine kinase. RET/PTC rearrangements are reported in sporadic PTC and postulated to be important genetic alterations in thyroid tumorigenesis (79-84). More than 10 RET/PTC chimeric rearrangements have been described in sporadic PTC (7, 10, 13, 16, 84). The reported frequency in PTC is widely distributed from 10% to 40% in adult patients (7-16, 84). A higher frequency (60%-70%) of RET rearrangements, particularly RET/PTC3, has been reported in pediatric patients with radiation history (84). A relatively high incidence of RET/PTC rearrangements has also been reported in early-stage small PTCs. These findings suggest that this genetic alteration occurs as an early event in PTC tumorigenesis (82, 83). The absence of RET/PTC rearrangements in most PDC and UC also suggests that RET/PTC rearrangements have a minor role in tumor progression, which means a lower risk of anaplastic transformation in PTC with RET/PTC rearrangements (7, 17, 65, 67, 68).

RET/PTC rearrangement, and *RAS* and *BRAF* mutations collectively account for 60%-70% of PTC and appear to be mutually exclusive. Kimura et al. emphasized the pivotal role of this kinase cascade in PTC development (65). However, Nakazawa et al. suggested that the *RET/PTC* rearrangement in PTC is not a driver mutation because the PET/PTC signal was found in only a minor population of tumor cells using *in situ* hybridization on cytological smear samples (86). Furthermore, *RET/PTC* rearrangement was found in benign thyroid nodule and their exact roles in thyroid carcinogenesis become unclear (87-89).

TRK rearrangements involve another receptor tyrosine kinase gene, *NTRK1* (1q22), and these rearrangements have been reported in up to 10%-15% of PTCs (90-93). Musholt et al. reported 12.6% of *TRK* rearrangements in 199 PTCs and the PTC with *TRK* rearrangement is associated with a higher (60%) local recurrence and a higher (27%) tumor-related mortality rate than in the PTC without *TRK* rearrangements (93).

c. Molecular Genetics of Poorly Differentiated Carcinoma and Undifferentiated Carcinoma

UC is a rare and highly aggressive thyroid carcinoma, with a mean survival of less than 6 months after diagnosis and PDC is an aggressive thyroid carcinoma with its morphological and behavioral features between WDC and UC, and often shows anaplastic transformation (7, 8, 11, 12, 16, 21, 36, 44, 94, 95). In the multistep carcinogenesis theory, UC is postulated to be derived from preexisting low-grade WDCs through the gain of additional mutations, such as loss of TP53 and further loss of tumor suppressors (7, 8, 11, 12, 16, 21, 36, 44, 68, 94-97). Soares et al. screened for and found BRAF mutations in 6/17 (35%) cases of UC and 1/3 UC-derived cell line, but none in insular-type PDC (68). They concluded that UC may progress from BRAF(V600E) mutated PTC and insular-type PDCs are more closely related to FTC than PTC (11, 68). There are many studies analyzing the genetic alterations and molecular profiles of UC, the end step in the thyroid multistep carcinogenesis; many of them conclude that BRAF-mutated UCs are often associated with BRAF-mutated PTC as a precursor lesion and progression from PTC to UC could be favored by further TP53 mutation and SOX2 expression, entering the final stage of progression of UC (7, 11-16, 41, 66, 69, 96). The TP53 tumor suppressor gene is located on chromosome 17p13 and encodes a nuclear transcription factor related to the cell cycle, DNA repair and apoptosis. TP53 point mutations are almost exclusively found in PDC and UC (17%-38% of PDC and 60%-88% of UC) and are regarded as a late event in the multistep carcinogenesis theory in contrast to that they are observed in the early phase of colorectal carcinogenesis (Table 2) (4-6, 11-13, 16, 37-45). Its deadly biological behavior and high proliferation rate (Ki-67 labeling index more than 30%), as well as genetic instability and marked aneuploidy, are attributed to accumulation of these genetic and epigenetic alterations (7, 98-100).

β-catenin is a cytoplasmic protein encoded by the *CTNNB1* gene located on chromosome 3p22-3p21.3. Point mutations in the *CTNNB1* gene have been reported in 25% of PDC and 66% of UC (Table 2), but none in WDCs (PTC and FTC) (103). Constitutive activation of the Wnt-signaling pathway by mutated *β-catenin* may play an important role in the progression from WDCs to PDC and UC (11, 103-106).

Murugan AK and Xing M reported two novel gain-offunction mutations of the *ALK* gene in 11% of UCs but no mutations in WDCs (107). Combination of *PI3K/Akt* genetic alterations (*PTEN*, *RAS*, *ALK*) with a *BRAF* mutation has been shown to be present in more aggressive thyroid tumors such as UC, and genetic or epigenetic alterations that activate both MAPK and PI3K/Akt pathways have been shown to be present in most cases of UC (42, 75, 108, 109). Other genes and pathways important for the development of PDC and UC may include the *LRP1B* gene located at 2q21, a susceptibility locus for familial non-medullary thyroid carcinomas (110). Hypermethylation of many tumor suppressor genes and multiple allelic losses, which may be linked to the inactivation of tumor suppressor genes, have been reported in thyroid carcinomas with poor outcome (30-32, 108, 110-121).

3. Molecular Changes in the Progression of Thyroid Carcinomas

The progression of thyroid carcinomas after initiation and promotion involves complex genetic and epigenetic alterations and mutations, and the epigenetic alterations in PDC and UC are far from being completely clarified (110-119). Growth factors (FGF, FGFR, MET, EGFR and VGEF) involved in the PI3K-Akt-mTOR pathway have been reported to have an important role in tumor development and progression in thyroid follicular cell carcinogenesis (15, 27, 33, 42, 109, 119-127). Liu et al. found copy number gains in EGFR, PDGFR (α and β), VEGFR1, VGEFR2, KIT, MET, PIK3 (a and b) and PDK1 genes, which may play important roles in the tumor progression to UC (42). Cell cycle regulators such as retinoblastoma (Rb), p21, p27 and TP53 also play important roles in the progression of thyroid carcinogenesis (49, 100, 126, 127). Adhesion molecules (β-catenin and E-cadherin) induce signal transduction through the Wnt signal pathway. Decreased E-cadherin expression is often found in UC, which activates β -catenin transcription activity by increasing the pool of β -catenin to migrate to the nucleus. Fibronectin is another adhesion molecule involved in the progression of thyroid carcinoma and the up-regulation of fibronectin has been demonstrated in thyroid carcinomas (33, 128).

4. Thyroid Carcinomas with Metastasis

Metastatic disease is the most common and important immediate cause of death in patients with thyroid carcinomas, either WDC or UC (21, 36, 44, 94, 126, 127). From the multistep carcinogenesis theory, FTC with metastasis should have some additional genetic alterations compared with those without metastasis, and this should also be the case for PTC. These genetic events in either PTC or FTC are far from being completely clarified. *N-RAS* codon 61 mutation has been reported to be an important predictive genetic alteration for distant metastasis in PTC (128) and FTC (49, 50, 54, 55). However, these genetic changes are observed in cases without invasion or metastasis, and the genetic alterations that specifically linked to invasion and metastasis have not been fully elucidated in thyroid carcinomas. Many studies have postulated that accumulation of the genetic and epigenetic alterations rather than a single genetic event are responsible for tumor progression from non-invasive to invasive or from non-metastasizing to metastasizing (15). A common hypermethylation of hMLH1 (DNA repair gene) has been reported to be associated with lymph node metastasis in PTC with BRAF mutation (113). BRAF(V600E) mutation has been reported to promote the progression and aggressiveness of PTC by downregulation of tumor suppressor genes and up-regulation of cancerpromoting molecules (VEGF, matrix metalloproteinases, nuclear transcription factor kappa B and c-met) (112). VEGF-D has been reported to play an important role in lymph node metastasis via lymphangiogenesis in PTC, and Nakamura Y et al. showed that upregulation of VEGF-D expression by nitric oxide in a cultured PTC cell line and the high level of nitrotyrosine (a biomarker for peroxynitrate formation from nitric oxide) immune expression was significantly correlated with lymph node metastasis (121, 129). Zuo M et al. showed that S100A4 was one of the overexpressed genes in a human thyroid carcinoma clone with a high incidence of lung metastasis using microarray analysis of gene expression profiling in their animal transplantation model (130). Zuo M et al. further reported a significantly higher level of S100A4 transcript in metastatic tumors than in the primary tumors (131). There are several genes (Nm23-H1, KiSS-1, RCAN1 and KAI1) that act as suppressors in tumor metastasis, and Phay JE et al. concluded that the loss of expression or function of such metastasis suppressor genes may play an important role in the development of metastasis (15). Since distant metastasis is one of the most important prognostic factors in thyroid carcinomas, identification of those genetic and epigenetic events could reveal new molecular targets for treating advanced thyroid carcinomas (126, 127).



Figure 1: Thyroid neoplasia and multistep carcinogenesis theory. PTC lineage: 1: unknown, 2: unknown, 3: *RET* rearrangement or *BRAF* mutation, 4: unknown, 5: cyclin D1 overexpression and p27 down regulation, 6: *TP53*, *PTEN* and *CTNNB1* mutation, 7: *TP53*, *PTEN*, *ALK*, *PIK3CA*, *LRP1B* and *CTNNB1* mutation.

FTC lineage: 1: unknown, 2: *RAS* mutation or *PPARG* rearrangement, 3: RAS mutation or PPARG rearrangement, 4: RAS mutation or *PPARG* rearrangement, 5: *N-RAS* codon 61 mutation, 6: *TP53*, *PTEN* and *CTNNB1* mutation, 7: *TP53*, *PTEN*, *ALK*, *PIK3CA*, *LRP1B* and *CTNNB1* mutation.

5. Multistep Carcinogenesis Theory

A simplified model of the multistep carcinogenesis theory is shown in Figure 1 and the known genetic changes of thyroid carcinomas discussed in the present review are incorporated into the figure legends (Fig. 1). Thyroid tumors develop as a result of the sequential accumulation of genetic and epigenetic alterations involved in the control of cell proliferation, cell differentiation or cell death, according to the multistep carcinogenesis theory. Additional epigenetic alterations also superimpose on those genetic changes in this model (15, 16, 109-117). One of the difficulties in this model is the lack of a clear cut separation of all histological types of thyroid carcinomas into corresponding tumor stages (stages 0, I, II, III and IV, or early non-invasive neoplasm, invasive carcinoma confined to thyroid gland, invasive carcinoma with local metastasis and invasive carcinoma with distant metastasis). This is because the age of patients is an important factor in the staging of thyroid carcinomas in most clinical guidelines and all patients younger than 45 years old are classified into stage I regardless of invasion and metastasis, and all UC patients are classified into stage IV even without invasion and metastasis (21, 36, 132). As a result, most of the literature on thyroid carcinogenesis handled thyroid carcinomas in histologic groups, such as PTC group, FTC group, PDC group and UC group, rather than the stage of tumors. Therefore, in this review, multistep theory and related genetic alterations are explained separately in the PTC lineage and the FTC lineage in the figure legends (Fig. 1). Other problems for understanding thyroid carcinogenesis are 1) socalled mutation-negative thyroid carcinomas (about one-third of PTC and FTC could not be explained by known genetic mutations) and 2) TP53 mutation is restricted in PDC and UC as a late genetic event in thyroid carcinogenesis, in contrast to the TP53 mutation in colorectal cancer (it occurs at an early phase of progression from adenoma to carcinoma in colorectal carcinoma) (4-6, 11). There are still many unanswered questions in this multistep theory when it is applied to thyroid follicular cell carcinogenesis.

6. A Missing Link in Multistep Carcinogenesis

There are several points that do not fit the multistep carcinogenesis theory in thyroid follicular cell neoplasms, and they are discussed in detail by Takano et al. elsewhere in this special issue of the Journal of Basic & Clinical Medicine (3). Another problem in explaining thyroid neoplasms using the multistep carcinogenesis model is a lack of known precursor lesions of thyroid carcinomas between normal follicular cells and early WDCs, a missing link in the multistep carcinogenesis of thyroid follicular cell carcinogenesis. Although FA is described as a candidate precursor lesion for thyroid carcinomas in many reviews and textbooks, prophylactic surgery is not recommended for patients with FA owing to infrequent progression from benign FA to malignant WDC (7, 8, 10, 13-16, 21, 51, 132-137). Furthermore, from many studies on how to separate malignant lesions (FTC and PTC) from FA using genetic markers, they act as if they are distinct lesions rather than a lesion with genetic continuity (134-143). In the PTC chapter of WHO blue book, it is explained that there is no known precursor lesion for PTC, and in the FTC chapter, it is stated that no definite precursor lesions of FTC are known (36). The possible precursor lesions of thyroid carcinomas, such as small papillary carcinoma and borderline lesions between normal follicular cells and WDCs including FA have been reviewed elsewhere (144-146).

References:

 Takano T. Fetal cell carcinogenesis of the thyroid: a hypothesis for better understanding of gene expression profile and genomic alteration in thyroid carcinoma. Endocr J 2004; 51:509-15.

- Takano T, Amino N. Fetal cell carcinogenesis: a new hypothesis for better understanding of thyroid carcinoma. Thyroid 2005; 15:432-8.
- 3. Takano T: The basic theory of fetal cell carcinogenesis of the thyroid. J Basic Clin Med 2014; 3:6-11.
- Vogelstein B, Fearon ER, Hamilton SR, Kern SE, Preisinger AC, Leppert M, Nakamura Y, White R, Smits AM, Bos JL. Genetic alterations during colorectal-tumor development. N Engl J Med 1988; 319:525-32.
- Vogelstein B, Fearon ER, Kern SE, Hamilton SR, Preisinger AC, Nakamura Y, White R. Allelotype of colorectal carcinomas. Science 1989; 244:207-11.
- Barrett JC. Mechanisms of multistep carcinogenesis and carcinogen risk assessment. Environment Health Perspect 1993; 100:9-20.
- Kondo T, Ezzat S, Asa SL. Pathogenetic mechanisms in thyroid follicular-cell neoplasia. Nat Rev Cancer 2006; 6:292-306.
- Nikiforov YE. Thyroid carcinoma: molecular pathways and therapeutic targets. Mod Pathol 2008; 21:S37-43.
- Eberhardt NL, Grebe SK, McIver B, Reddi HV. The role of the PAX8/PPAR gamma fusion oncogene in the pathogenesis of follicular thyroid cancer. Mol Cell Endocrinol 2010; 321:50-6.
- Parameswaran R, Brooks S, Sadler GP. Molecular pathogenesis of follicular cell derived thyroid cancers. Int J Surg 2010; 8:186-93.
- Soares P, Lima J, Preto A, Castro P, Vinagre J, Celestino R, Couto JP, Prazeres H, Eloy C, Máximo V, Sobrinho-Simões M. Genetic alterations in poorly differentiated and undifferentiated thyroid carcinomas. Curr genomics 2011; 12:609-17.
- Gauchotte G, Philippe C, Lacomme S, Leotard B, Wissler MP, Allou L, Toussaint B, Klein M, Vignaud JM, Bressenot A. BRAF, p53 and SOX2 in anaplastic thyroid carcinoma: evidence for multistep carcinogenesis. Pathology 2011; 43:447-52.
- Nikiforov YE, Nikiforova MN. Molecular genetics and diagnosis of thyroid cancer. Nat Rev Endocrinol 2011; 30:569-80.
- 14. Xing M. Molecular pathogenesis and mechanisms of thyroid cancer. Nat Rev Cancer 2013; 13:184-99.
- Phay JE, Ringel MD. Metastatic mechanisms in follicular cell-derived thyroid cancer. Endocr Relat Cancer 2013; 20:R307-R319.
- Omur O, Baran Y. An update on molecular biology of thyroid cancers. Crit Rev Oncol Hematol 2014; 90:233-52.
- Ozaki O, Ito K, Kobayashi K, Suzuki A, Manabe Y, Hosoda Y. Familial occurrence of differentiated, non-medullary thyroid carcinoma. World J Surg 1988; 12:565-71.
- Goldgar DE, Easton DF, Cannon-Albright LA, Skolnick MH. Systematic population-based assessment of cancer risk in first-degree relatives of cancer probands. J Natl Cancer Inst 1994; 86:1600-8.
- Nose V. Familial non-medullary thyroid carcinoma: an update. Endocr Pathol 2008; 19:226-40.
- Ito Y, Kakudo K, Hirokawa M, Fukushima M, Yabuta T, Tomoda C, Inoue H, Kihara M, Higashiyama T, Uruno T, Takamura Y, Miya A, Kobayashi K, Matsuzuka F, Miyauchi A. Biological behavior and prognosis of familial papillary thyroid carcinoma. Surgery 2009; 145:100-5.
- 21. Kahrn A, Nose V. Pathology of thyroid gland in Endocrine pathology, Differential diagnosis and molecular advances,

second edition, editor Lloyd RV, Springer, New York, 2010; p181-236.

- 22. Fallah M, Pukkala E, Tryggvadottir L, Olsen JH, Tretli S, Sundquist K, Hemminki K. Risk of thyroid cancer in firstdegree relatives of patients with non-medullary thyroid cancer by histology type and age at diagnosis: a joint study from five Nordic countries. J Med Genet 2013; 50:373-82.
- Halachmi N, Harachimi S, Evron E, Cairns P, Okami K, Saji M, Westra WH, Zeiger MA, Jen J, Sidransky D. Somatic mutations of the PTEN tumor suppressor gene in sporadic follicular thyroid tumors. Genes Chromosomes Cancer 1998; 23:239-43.
- 24. Gimm O, Perren A, Weng LP, Marsh DJ, Yeh JJ, Ziebold U, Gil E, Hinze R, Delbridge L, Lees JA, Mutter GL, Robinson BG, Komminoth P, Dralle H, Eng C. Differential nuclear and cytoplasmic expression of PTEN in normal thyroid tissue, and benign and malignant epithelial thyroid tumors. Am J Pathol 2000; 156:1693-700.
- Eng C. Role of PTEN, a lipid phosphatase upstream effector of protein kinase B, in epithelial thyroid carcinogenesis. Ann N Y Acad Sci 2002; 968:213-21.
- Alvarez-Nunez F, Bussaglia E, Mauricio D, Ybarra J, Vilar M, Lerma E, de Leiva A, Matias-Guiu X; Thyroid Neoplasia Study Group. PTEN promoter methylation in sporadic thyroid carcinoma. Thyroid 2006; 16:17-23.
- Saji M, Ringel MD. The PI3K-Akt-mTOR pathway in initiation and progression of thyroid tumors. Mol Cell Endocrinol 2010; 321:20-8.
- Soares P, Berx G, van Roy F, Sobrinho-Simões M. Ecadherion gene alterations are rare events in thyrod tumors. Int J Cancer 1997; 70:32-8.
- Hemmer S, Wasenius VM, Knuutila S, Franssila K, Joensuu H. DNA copy number changes in thyroid carcinoma. Am J Pathol 1999; 154:1539-47.
- Kitamura Y, Shimizu K, Tanaka S, Ito K, Emi M. Allelotyping of anaplastic thyroid carcinoma: frequent allelic losses on 1q, 9p, 11, 17, 19p, and 22q. Genes Chromosomes Cancer 2000; 27:244-51.
- 31. Kitamura Y, Shimizu K, Tanaka S, Ito K, Emi M. Association of allelic loss on 1q, 4p, 7q, 9p, 9q, and 16q with postoperative death in papillary thyroid carcinoma. Clin Cancer Res 2000; 6:1819-25.
- 32. Kitamura Y, Shimizu K, Ito K, Tanaka S, Emi M. Allelotyping of follicular thyroid carcinoma: frequent allelic losses in chromosome arms 7q, 11p, and 22q. J Clin Endocrinol Metab 2001; 86:4268-72.
- 33. Huang Y, Prasad M, Lemon WJ, Hampel H, Wright FA, Kornacker K, LiVolsi V, Frankel W, Kloos RT, Eng C, Pellegata NS, de la Chapelle A. Gene expression in papillary thyroid carcinoma reveals highly consistent profiles. Proc Natl Acad Sci USA 2001; 98:15044-9.
- Rodrigues RF, Roque L, Rosa-Santos J, Cid O, Soares J. Chromosomal imbalance associated with anaplastic transformation of follicular thyroid carcinomas. Br J Cancer 2004; 90:492-6.
- 35. Lee J, Hwang JA, Lee EK. Recent progress of genome study for anaplastic thyroid cancer. Genomics inform 2013; 11:68-75.
- DeLellis RA, Lloyd RV, Heitz PU, Eng C. Tumors of Endocrine Organs, World Health Organization Classification of Tumors; Pathology and Genetics. Lyon: IARC Press; 2004.
- 37. Ito T, Seyama T, Mizuno T, Tsuyama N, Hayashi T, Hayashi Y, Dohi K, Nakamura N, Akiyama M. Unique association of p53

mutations with undifferentiated but not with differentiated carcinomas of the thyroid gland. Cancer Res 1992; 52:1369-71.

- Fagin JA, Matsuo K, Karmakar A, Chen DL, Tang SH, Koeffler HP. High prevalence of mutations of the p53 gene in poorly differentiated human thyroid carcinomas. J Clin Invest 1993; 91:179-84.
- Donghi R, Longoni A, Pilotti S, Michieli P, Della Porta G, Pierotti MA. Gene p53 mutations are restricted to poorly differentiated and undifferentiated carcinomas of the thyroid gland. J Clin Invest 1993; 91:1753-60.
- 40. Garcia-Rostan G, Zhao H, Campo RL, Pollan M, Herrero A, Pardo J, Wu R, Carcangiu ML, Costa J, Tallini G Ras mutations are associated with aggressive tumor phenotypes and poor prognosis in thyroid cancer. J Clin Oncol 2003; 21:3226-35.
- 41. Quiros RM, Ding HG, Gattuso P, Prinz RA, Xu X. Evidence that one subset of anaplastic thyroid carcinomas are derived from papillary carcinomas due to BRAF and p53 mutations. Cancer 2005; 103: 2261-8.
- 42. Liu Z, Hou P, Ji M, Guan H, Studeman K, Jensen K, Vasko V, El-Naggar AK, Xing M. Highly prevalent genetic alterations in receptor tyrosine kinases and phosphatidylinositol 3kinase/akt and mitogen-activated protein kinase pathways in anaplastic and follicular thyroid cancers. J Clin Endocrinol Metab 2008; 93:3106-16.
- Pita JM, Banito A, Cavaco BM, Leite V. Gene expression profiling associated with the progression to poorly differentiated thyroid carcinomas. Br J Cancer 2009; 101:1782-91.
- O'Neill JP, Shaha AR. Anaplastic thyroid cancer. Oral Oncol 2013; 49:702-6.
- 45. Pita, JM, Figueiredo IF, Moura MM, Leite V, Cavaco BM. Cell cycle regulation and TP53 and RAS mutations are major events in poorly differentiated and undifferentiated thyroid carcinomas. J Clin Endocrinol Metab 2014; 99:E495-507.
- Suarez HG, Du Villard JA, Caillou B, Schlumberger M, Tubiana M, Parmentier C, Monier R. Detection of activated ras oncogenes in human thyroid carcinomas. Oncogene 1988; 2:403-6.
- Lemoine NR, Mayall ES, Wyllie ES, Williams ED, Goyns M, Stringer B, Wynford-Thomas D. High frequency of ras oncogene activation in all stage of human thyroid tumorigenesis. Oncogene 1989; 4:159-64.
- Namba H, Rubin SA, Faqin JA. Point mutations of ras oncogenes are an early event in thyroid tumorigenesis. Mol Endocrinol 1990; 4:1474-9.
- 49. Basolo F, Pisaturo F, Pollina LE, Fontanini G, Elisei R, Molinaro E, Iacconi P, Miccoli P, Pacini F. NRAS mutation in poorly differentiated thyroid carcinoma: correlation with bone metastases and inverse correlation to thyroglobulin expression. Thyroid 2000; 10:19-23.
- Motoi N, Sakamoto A, Yamochi T, Horiuchi H, Motoi T, Machinami R. Role of ras mutation in the progression of thyroid carcinoma of follicular epithelial origin. Pathol Res Pract 2000; 196:1-7.
- Nikiforova MN, Lynch RA, Biddinger PW, Alexander EK, Dorn GW 2nd, Tallini G, Kroll TG, Nikiforov YE. RAS point mutations and PAX8-PPAR gamma rearrangement in thyroid tumors: evidence for distinct molecular pathways in thyroid follicular carcinoma. J Clin Endocrinol Metab 2003; 88:2318-26.

- 52. Wang HM, Huang YW, Huang JS, Wang CH, Kok VC, Hung CM, Chen HM, Tzen CY. Anaplastic carcinoma of the thyroid arising more often from follicular carcinoma than papillary carcinoma. Ann Surg Oncol 2007; 14:3011-8.
- 53. Freitas BC, Cerutti JA. Genetic Markers differentiating follicular thyroid carcinoma from benign lesions. Mol Cell Endocrinol 2010; 321:77-85.
- 54. Fukahori M, Yoshida A, Hayashi H, Yoshihara M, Matsukuma S, Sakuma Y, Koizume S, Okamoto N, Kondo T, Masuda M, Miyagi Y. The association between RAS mutations and clinical characteristics in follicular thyroid tumors: new insights from a single center and a large patient cohort. Thyroid 2012; 22:683-9.
- 55. Jang EK, Song DE, Sim SY, Kwon H, Choi YM, Jeon MJ, Han JM, Kim WG, Kim TY, Shong YK, Kim WB. *NRAS* codon 61 mutation is associated with distant metastasis in patients with follicular thyroid carcinoma. Thyroid 2014; 24:1275-81.
- Kroll TG, Sarraf P, Pecciarini L, Chen CJ, Mueller E, Spiegelman BM, Fletcher JA. PAX8-PPAR gamma1 fusion oncogene in human thyroid carcinoma. Science 2000; 289:1357-60.
- Nikiforova MN, Biddinger PW, Caudill CM, Kroll TG, Nikiforov YE. PAX8-PPARgamma rearrangement in thyroid tumors: RT-PCR and immunohistochemical analysis. Am J Surg Pathol 2002; 26:101601023.
- Marques AR, Espandinha C, Catarino AL, Moniz S, Pereira T, Sobrinho LG, Leite V. Expression of PAX8-PPAR gamma 1 rearrangements in both follicular thyroid carcinoma and adenomas. J Clin Endocrinol Metab 2002; 87:3947-52.
- 59. Kakudo K, Bai Y, Katayama S, Hirokawa M, Ito Y, Miyauchi A, Kuma K. Classification of follicular cell tumors of the thyroid gland: analysis involving Japanese patients from one institute. Pathol Int 2009; 59:359-67.
- Fukushima T, Suzuki S, Mashiko M, Ohtake T, Endo Y, Takebayashi Y, Sekikawa K, Hagiwara K, Takenoshita S. BRAF mutations in papillary carcinomas of the thyroid. Oncogene 2003; 22: 6455-67.
- Namba H, Nakashima M, Hayashi T, Hayashida N, Maeda S, Rogounovitch TI, Ohtsuru A, Saenko VA, Kanematsu T, Yamashita S. Clinical implication of hot spot BRAF mutation, V599E, in papillary thyroid cancers. J Clin Endocrinol Metab 2003; 88:4393-7.
- 62. Soares P, Trovisco V, Rocha AS, Lima J, Castro P, Preto A, Máximo V, Botelho T, Seruca R, Sobrinho-Simões M. BRAF mutations and RET/PTC rearrangements are alternative events in the etiopathogenesis of PTC. Oncogene 2003; 17:4578-80.
- Cohen Y, Xing M, Mambo E, Guo Z, Wu G, Trink B, Beller U, Westra WH, Ladenson PW, Sidransky D. BRAF mutation in papillary thyroid carcinoma. J Natl Cancer Inst 2003; 95:625-7.
- 64. Xu X, Quiros RM, Gattuso P, Ain KB, Prinz RA. High prevalence of BRAF gene mutation in papillary thyroid carcinomas and thyroid tumor cell lines. Cancer Res 2003; 63:4561-7.
- 65. Kimura ET, Nikiforova MN, Zhu Z, Knauf JA, Nikiforov YE, Fagin JA. High prevalence of BRAF mutations in thyroid cancer: genetic evidence for constitutive activation of the RET/PTC-RAS-BRAF signaling pathway in papillary thyroid carcinoma. Cancer Res 2003; 63:1454-7.
- 66. Nikiforova, MN, Kimura ET, Gandhi M, Biddinger PW, Knauf JA, Basolo F, Zhu Z, Giannini R, Salvatore G, Fusco A,

Santoro M, Fagin JA, Nikiforov YE. BRAF mutations in thyroid tumors are restricted to papillary carcinomas and anaplastic or poorly differentiated carcinomas arising from papillary carcinomas. J Clin Endocrinol Metab 2003; 88:5399-404.

- 67. Trovisco V, Vieira de Castro I, Soares P, Máximo V, Silva P, Magalhães J, Abrosimov A, Guiu XM, Sobrinho-Simões M. BRAF mutations are associated with some histological types of papillary thyroid carcinoma. J Pathol 2004; 202:247-51.
- 68. Soares P, Trovisco V, Rocha AS, Feijão T, Rebocho AP, Fonseca E, Vieira de Castro I, Cameselle-Teijeiro J, Cardoso-Oliveira M, Sobrinho-Simões M. BRAF mutations typical of papillary thyroid carcinoma are more frequently detected in undifferentiated than in insular and insular-like poorly differentiated carcinomas. Virch Arch 2004; 444:572-6.
- 69. Kim J, Giuliano AE, Turner RR, Gaffney RE, Umetani N, Kitago M, Elashoff D, Hoon DS. Lymphatic mapping establish the role of BRAF gene mutation in papillary thyroid carcinoma. Ann Surg 2006; 244:799-804.
- Ugolini C, Giannini R, Lupi C, Salvatore G, Miccoli P, Proietti A, Elisei R, Santoro M, Basolo F. Presence of BRAFV600E in very early stages of papillary thyroid carcinoma. Thyroid 2007; 17:381-8.
- 71. Li Y, Nakamura M, Kakudo K. Targeting of the BRAF gene in papillary thyroid carcinoma (review). Oncol Rep 2009; 22:671-81.
- Xing M. BRAF mutation in papillary thyroid cancer. pathogenic role, molecular bases, and clinical implications. Endocr Rev 2007; 28:742-62.
- Lupi C, Giannini R, Ugolini C, Proietti A, Berti P, Minuto M, Materazzi G, Elisei R, Santoro M, Miccoli P, Basolo F. Association of BRAFV600E mutation with poor clinicopathological outcomes in 500 consecutive cases of papillary thyroid carcinoma. J Clin Endocrinol Metab 2007; 92:4085-90.
- 74. Ricarte-Filho JC, Ryder M, Chitale DA, Rivera M, Heguy A, Ladanyi M, Janakiraman M, Solit D, Knauf JA, Tuttle RM, Ghossein RA, Fagin JA. Mutational profile of advanced primary and metastatic radioactive iodine-refractory thyroid cancers reveals distinct pathogenetic roles for *BRAF*, *PIK3CA* and *AKT1*. Cancer Res 2009; 69:4885-93.
- 75. Xing M, Alzahrani AS, Carson KA, Viola D, Elisei R, Bendlova B, Yip L, Mian C, Vianello F, Tuttle RM, Robenshtok E, Fagin JA, Puxeddu E, Fugazzola L, Czarniecka A, Jarzab B, O'Neill CJ, Sywak MS, Lam AK, Riesco-Eizaguirre G, Santisteban P, Nakayama H, Tufano RP, Pai SI, Zeiger MA, Westra WH, Clark DP, Clifton-Bligh R, Sidransky D, Ladenson PW, Sykorova V. Association between *BRAF* V600E mutation and mortality in patients with papillary thyroid cancer. JAMA 2013; 309:1493-501.
- 76. Ito Y, Yoshida H, Maruo R, Morita S, Takano T, Hirokawa M, Yabuta T, Fukushima M, Inoue H, Tomoda C, Kihara M, Uruno T, Higashiyama T, Takamura Y, Miya A, Kobayashi K, Matsuzuka F, Miyauchi A. BRAF mutation in papillary thyroid carcinoma in Japanese population: its lack of correlation with high-risk clinicopathological features and disease-free survival of patients. Endocr J 2009; 56:89-97.
- 77. Eloy C, Santos J, Soares P, Sobrinho-Simões M. The preeminence of growth pattern and invasiveness and the limited influence of BRAF and RAS mutations in the occurrence of papillary carcinoma lymph node metastases. Virch Arch 2011; 459:265-76.
- 78. Guerra A, Sapio MR, Marotta V, Campanile E, Rossi S, Forno

I, Fugazzola L, Budillon A, Moccia T, Fenzi G, Vitale M. The primary occurrence of BRAFV600E is a rare clonal event in papillary thyroid carcinoma. J Clinical Endocrinol Metab 2012; 97:517-24.

- 79. Ishizaka Y, Itoh F, Tahira T, Ikeda I, Ogura T, Sugimura T, Nagao M. Presence of aberrant trascripts of ret protooncogene in a human papillary thyroid carcinoma cell line. Jpn J Cancer Res 1989; 80:1149-52.
- 80. Grieco M, Santoro M, Berlingieri MT, Melillo RM, Donghi R, Bongarzone I, Pierotti MA, Della Porta G, Fusco A, Vecchio G. PTC is a novel rearranged form of the ret proto-oncogene and is frequently detected in vivo in human thyroid papillary carcinomas. Cell 1990; 60:557-63.
- Santro M, Carlomagno F, Hay ID, Herrmann MA, Grieco M, Melillo R, Pierotti MA, Bongarzone I, Della Porta G, Berger N. Ret oncogene activation in human thyroid neoplasms is restricted to the papillary cancer subtype. J Clin Invest 1992; 89:17-22.
- Viglietto G, Chiappetta G, Martinez-Tello FJ, Fukunaga FH, Tallini G, Rigopoulou D, Visconti R, Mastro A, Santoro M, Fusco A. RET/PTC oncogene activation is an early event in thyroid carcinogenesis. Oncogene 1995; 11:1207-10.
- Sugg SL, Ezzat S, Rosen IB, Freeman JL, Asa SL. Distinct multiple RET/PTC gene rearrangements in multifocal papillary thyroid neoplasia. J Clin Endocrinol Metab 1998; 83:4116-22.
- Nikiforov YE, Rowland JM, Bove KE, Monforte-Munoz H, Fagin JA. Distinct pattern of ret oncogene rearrangements in morphological variants of radiation-induced and sporadic thyroid papillary carcinomas in children. Cancer Res 1997; 57:1690-4.
- Tallini G, Santro M, Helie M, Carlomagno F, Salvatore G, Chiappetta G, Carcangiu ML, Fusco A. RET/PTC oncogene activation defines a subset of papillary thyroid carcinomas lacking evidence of progression to poorly differentiated or undifferentiated tumor phenotypes. Clin Cancer Res 1998; 4:287-94.
- Kimura ET, Nikiforova MN, Zhu Z, Knauf JA, Nikiforov YE, Fagin JA. High prevalence of BRAF mutations in thyroid cancer. Genetic evidence for constitutive activation of the RET/PTC-RAS-BRAF signaling pathway in papillary thyroid carcinoma. Cancer Res 2003; 1931-40.
- 87. Nakazawa T, Murata S, Kondo T, Niu D, Mochizuki K, Kawasaki T, Yamane T, Nakamura N, Katoh R. RET/PTC rearrangements arising from a small population of papillary thyroid carcinoma cells possible candidate for passenger mutation. Virch Arch 2009; 455:35-41.
- Ishizaka Y, Kobayashi S, Ushijima T, Hirohashi S, Sugimura T, Nagao M. Detection of retTPC/PTC transcripts in thyroid adenomas and adenomatous goiter by an RT-PCR method. Oncogene 1991; 6:1667-72.
- Guerra A, Sapio MR, Marotta V, Campanile E, Moretti MI, Deandrea M, Motta M, Limone PP, Fenzi G, Rossi G, Vitale M. Prevalence of RET/PTC rearrangement in benign and malignant thyroid nodules and its clinical application. Endocr J 2011; 58:31-8.
- 90. Sapio MR, Guerra A, Marotta V, Campanile E, Formisano R, Deandrea M, Motta M, Limone PP, Fenzi G, Rossi G, Vitale M. High growth rate of benign thyroid nodules bearing RET/PTC rearrangements. J Clin Endocrinol Metab 2011; 96:E916-9.
- Pierotti MA, Bongarzone I, Borrello MG, Mariani C, Miranda C, Sozzi G, Greco A. Rearrangement of TRK proto-oncogene in papillary thyroid carcinomas. J Endocrinol Invest 1995;

18:130-3.

- 92. Pierotti MA, Bongarzone I, Borello MG, Greco A, Pilotti S, Sozzi G. Cytogenetics and molecular genetics of the carcinomas arising from the thyroid epithelial follicular cells. Genes Chromosomes Cancer 1996; 16:1-14.
- Musholt TJ, Musholt PB, Khaladj N, Schulz D, Scheumann GF, Klempnauer J. Prognostic significance of RET and NTRK1 rearrangements in sporadic papillary thyroid carcinoma. Surgery 2000; 128:984-93.
- Greco A, Miranda C, Pierotti MA. Rearrangements of NTRK1 gene in papillary thyroid carcinoma. Mol Cell Endocrinol 2010; 321:44-9.
- 95. Smallridge RC, Marlow LA, Copland JA. Anaplastic thyroid cancer: molecular pathogenesis and emerging therapies. Endocr Relat Cancer 2008; 16:17-44.
- Sugitani I, Miyauchi A, Sugino K, Okamoto T, Yoshida A, Suzuki S. Prognostic factors and treatment outcomes from anaplastic thyroid carcinoma: ATC Research Consortium of Japan cohort study of 677 patients. World J Surg 2012; 36:1247-54.
- 97. Mochizuki K, Kondo T, Nakazawa T, Iwashina M, Kawasaki T, Nakamura N, Yamane T, Murata S, Ito K, Kameyama K, Kobayashi M, Katoh R. RET rearrangements and BRAF mutation in undifferentiated thyroid carcinomas having papillary carcinoma components. Histopathol 2010; 57:444-50.
- Sarlis NJ. Expression pattern of cellular growth-controlling genes in non-medullary thyroid cancer: basic aspect. Rev Endocr Metab Disord 2000; 1:183-96.
- Erickson LA, Jin L, Wollan PC, Thompson GB, van Heerden J, Lloyd RV. Expression of p27kip1 and Ki-67 in benign and malignant thyroid tumors. Mod Pathol 1998; 11:169-74.
- 100. Wreesmann VB, Ghossein RA, Patel SG, Harris CP, Schnaser EA, Shaha AR, Tuttle RM, Shah JP, Rao PH, Singh B. Genomewide appraisal of thyroid cancer progression. Am J Pathol 2002; 161:1549-56.
- 101. Kakudo K, Wakasa T, Ohta Y, Yane K, Ito Y, Yamashita H. Prognostic classification of thyroid follicular cell tumors using Ki-67 labeling index: Risk stratification of thyroid follicular cell carcinomas [review]. Endocr J 2015; 62:1-12.
- 102. Van Aken E, De Wever O, Correia da Rocha AS, Mareel M. Defective E-cadherin/catenin complexes in human cancer. Virch Arch 2001; 439:725-51.
- 103. Cerrato A, Fulciniti F, Avallone A, Benincasa G, Palombini L, Grieco M. Beta- and gamma-catenin expression in thyroid carcinomas. J Pathol 1998; 185:267-72.
- 104. Garcia-Rostan G, Camp RL, Herrero A, Carcangiu ML, Rimm DL, Tallini G. β-Catenin dysregulation in thyroid neoplasms. Down-regulation, aberrant nuclear expression, and CTNNB1 exon 3 mutations are markers for aggressive tumor phenotype and poor prognosis. Am J Pathol 2001; 158:987-96.
- 105. Sastre-Perona A, Santisteban P. Role of the Wnt pathway in thyroid carcinoma. Front Endocrinol 2012; 3:1-10.
- 106. Murugan AK, Xing M. Anaplastic thyroid cancers harbor novel oncogenic mutations of ALK gene. Cancer Res 2011; 71:4403-11.
- 107. Wu G, Mambo E, Guo Z, Hu S, Huang X, Gollin SM, Trink B, Ladenson PW, Sidransky D, Xing M. Uncommon mutation, but common amplifications, of the PIK3CA gene in thyroid tumors. J Clin Endocrinol Metab 2005; 90:4688-93.
- 108. Santarpia L, El-Naggar AK, Cote GJ, Myers JN, Sherman SI. Phosphatidylinositol 3-kinase/akt and ras/raf-mitogenactivated protein kinase pathway mutations. J Clin Endocrinol

Metab 2008; 93:278-84.

- 109. Prazeres H, Torres J, Rodrigues F, Pinto M, Pastoriza MC, Gomes D, Cameselle-Teijeiro J, Vidal A, Martins TC, Sobrinho-Simões M, Soares P. Chromosomal, epigenetic and micro RNA-mediated inactivation of LRP1B, a modulator of the extracellular environment of thyroid cancer cells. Oncogene 2011; 30:1302-17.
- 110. Schagdarsurengin U, Gimm O, Hoang-Vu C, Dralle H, Pfeifer GP, Dammann R. Frequent epigenetic silencing of the CpG island promoter of RASSF1A in thyroid carcinoma. Cancer Res 2002; 62:3698-701.
- 111. Schagdarsurengin U, Gimm O, Dralle H, Hoang-Vu C, Dammann R. CpG island methylation of tumor-related promoters occurs preferentially in undifferentiated carcinoma. Thyroid 2006; 16:633-42.
- 112. Hu S, Liu D, Tufano RP, Carson KA, Rosenbaum E, Cohen Y, Holt EH, Kiseljak-Vassiliades K, Rhoden KJ, Tolaney S, Condouris S, Tallini G, Westra WH, Umbricht CB, Zeiger MA, Califano JA, Vasko V, Xing M. Association of aberrant methylation of tumor suppressor genes with tumor aggressiveness and BRAF mutation in papillary thyroid cancer. Int J Cancer 2006; 119:2322-9.
- 113. Xing M: Gene methylation in thyroid tumorigenesis. Endocrinol 2007; 148:948-53.
- 114. Guan H, Ji M, Hou P, Liu Z, Wang C, Shan Z, Teng W, Xing M. Hypermethylation of DNA mismatch repair gene hMLH1 and its association with lymph node metastasis and T1799A BRAF mutation in patients with papillary thyroid carcinoma. Cancer 2008; 113:247-55.
- 115. Kondo T, Asa SL, Ezzat S. Epigenetic dysregulation in thyroid neoplasia. Endocrinol Metab North Am 2008; 37:389-400.
- 116. Kondo T, Nakazawa T, Ma D, Niu D, Mochizuki K, Kawasaki T, Nakamura N, Yamane T, Kobayashi M, Katoh R. Epigenetic silencing of TTF1/NKX2-1 through DNA hypermethylation and histone H3 modulation in thyroid carcinomas. Lab Invest 2009; 89:791-9.
- 117. Hou P, Liu D, Xing M. Genome-wide alterations in gene methylation by the BRAF V600E mutation in papillary thyroid cancer cells. Endocr Relat Cancer 2011; 687-97.
- 118. Manchikova V, Buj R, Castelblanco E, Inglada-Pérez L, Diez A, de Cubas AA, Curras-Freixes M, Maravall FX, Mauricio D, Matias-Guiu X, Puig-Domingo M, Capel I, Bella MR, Lerma E, Castella E, Reverter JL, Peinado MÁ, Jorda M, Robledo M. DNA methylation profiling of well-differentiated thyroid cancer uncovers markers of recurrence free survival. Int J Cancer 2014; 135:598-610.
- 119. Wreesmann VB, Ghossein RA, Patel SG, Harris CP, Schnaser EA, Shaha AR, Tuttle RM, Shah JP, Rao PH, Singh B. Genome-wide appraisal of thyroid cancer progression. Am J Pathol 2002; 161:1549-56.
- 120. Lee J, Hwang JA, Lee EK. Recent progress of genome study for anaplastic thyroid cancer. Genomics Inform 2013; 11:68-75.
- 121. Klein M, Vignaud JM, Hennequin V, Toussaint B, Bresler L, Plénat F, Leclère J, Duprez A, Weryha G Increased expression of the vascular endothelial growth factor is a preoperative prognostic marker in papillary thyroid carcinoma. J Clin Endocrinol Metab 2001; 86:656-8.
- 122. Lennard CM, Patel A, Wilson J, Reinhardt B, Tuman C, Fenton C, Blair E, Francis GL, Tuttle RM. Intensity of vascular endothelial growth factor expression is associated with increased risk of recurrence and disease-free survival in

papillary thyroid cancer. Surg 2001; 129:552-8.

- 123. Yasuoka H, Nakamura Y, Zuo H, Tang W, Takamura Y, Miyauchi A, Nakamura M, Mori I, Kakudo K. VEGF-D expression and lymph vessels play an important role for lymph node metastasis in papillary thyroid carcinoma. Mod Pathol 2005; 18:1127-33.
- 124. Kondo T, Zheng L, Liu W, Kurebayashi J, Asa SL, Ezzat S. Epigenetically controlled fibroblast growth factor receptor 2 signaling imposes on the RAS/BRAF/mitogen-activated protein kinase pathway to modulate thyroid cancer progression. Cancer Res 2007; 67(11):5461-70.
- 125. Holm R, Nesland JM. Retinoblastoma and p53 tumor suppressor gene protein expression in carcinoma of the thyroid gland. J Pathol 1994; 172:267-72.
- 126. Shi Y, Zou M, Farid NR, al-Sedairy ST. Evidence of gene deletion of p21 (WAF1/CIP1), a cyclin-dependent protein kinase inhibitor, in thyroid carcinomas. Br J cancer 1996; 74:1336-41.
- 127. Takano T, Matsuzuka F, Miyauchi A, Yokozawa T, Liu G, Morita S, Kuma K, Amino N. Restricted expression of oncofetal fibronectin mRNA in thyroid papillary and anaplastic carcinoma: an in situ hybridization study. Br J Cancer 1998; 78:221-224.
- 128. Kitamura Y, Shimizu K, Nagahama M, Sugino K, Ozaki O, Mimura T, Ito K, Ito K, Tanaka S. Immediate cause of death in thyroid carcinoma: clinicopathological analysis of 161 fatal cases. J Clin Endocrinol Metab 1999; 84:4043-9.
- 129. Nixon IJ, Whitcher MM, Palmer FL, Tuttle RM, Shaha AR, Shah JP, Patel SG, Ganly I. The impact of distant metastases at presentation on prognosis in patients with differentiated carcinoma of the thyroid gland. Thyroid 2012; 22:884-9.
- 130. Hara H, Fulton N, Yashiro T, Ito K, DeGroot LJ, Kaplan EL. N-ras mutation: an independent prognostic factor for aggressiveness of papillary thyroid carcinoma. Surg 1944; 116:1010-6.
- 131. Nakamura Y, Yasuoka H, Zuo H, Takamura Y, Miyauchi A, Nakamura M, Kakudo K. Nitric oxide in papillary thyroid carcinoma: induction of vascular endothelial growth factor-D and correlation with lymph node metastasis. J Clin Endocrinol Metab 2006; 91:1582-5.
- 132. Zuo M, Famulski KS, Parhar RS, Baitei E, Al-Mohanna FA, Farid NR, Shi Y. Microarray analysis of metastasis-associated gene expression profiling in a murine model of thyroid carcinoma pulmonary metastasis: identification of S100A4(Mts1) gene overexpression as a poor prognostic marker for thyroid carcinoma. J Clin Endocrinol Metab 2004; 89:6146-54.
- 133. Zuo M, Al-Baradie RS, Al-Hindi H, Farid NR, Shi Y. S100A4 (Mts1) gene overexpression is associated with invasion and metastasis of papillary thyroid carcinoma. Br J Cancer 2005; 93:1277-84.
- 134. Cooper DS, Doherty GM, Haugen BR, Kloos RT, Lee SL, Mandel SJ, Mazzaferri EL, McIver B, Pacini F, Schlumberger M, Sherman SI, Steward DL, Tuttle RM. Revised American Association management guidelines for patients with thyroid nodules and differentiated thyroid cancer. Thyroid 2009; 19:1167-214.
- 135. Kuma K, Matsuzuka F, Kobayashi A, Hirai K, Morita S, Miyauchi A, Katayama S, Sugawara M. Outcome of long standing solitary thyroid nodules. World J Surg 1992; 16:583-7.
- 136. Alexander EK, Hurwitz S, Heering JP, Benson CB, Frates MC, Doubilet PM, Cibas ES, Larsen PR, Marqusee E. Natural

history of benign solid and cystic thyroid nodules. Ann Intern Med 2003; 138:15-8.

- 137. Alexander EK, Kennedy GC, Baloch ZW, Cibas ES, Chudova D, Diggans J, Friedman L, Kloos RT, LiVolsi VA, Mandel SJ, Raab SS, Rosai J, Steward DL, Walsh PS, Wilde JI, Zeiger MA, Lanman RB, Haugen BR. Preoperative diagnosis of benign thyroid nodules with indeterminate cytology. N Engl J Med 2012; 367:705-15.
- 138. Lim DJ, Kim JY, Baek KH, Kim MK, Park WC, Lee JM, Kang MI, Cha BY. Natural course of cytologically benign thyroid nodules: observation of ultrasonographic changes. Endocrinol Metab 2013; 28:110-8.
- 139. Lee S, Skelton TS, Zheng F, Schwartz KA, Perrier ND, Lee JE, Bassett RL, Ahmed S, Krishnamurthy S, Busaidy NL, Grubbs EG. The biopsy-proven benign thyroid nodule: is long term follow-up necessary? J Am Coll Surg 2013; 217:81-8.
- 140. Ohori NP, Nikiforova MN, Schoedel KE, LeBeau SO, Hodak SP, Seethala RR, Carty SE, Ogilvie JB, Yip L, Nikiforov YE. Contribution of molecular testing to thyroid fine-needle aspiration cytology of "follicular lesion of undetermined significance/atypia of undetermined significance". Cancer Cytopathol 2010; 118:178-23.
- 141. Freitas BC, Cerutti JM. Genetic markers differentiating follicular thyroid carcinoma from benign lesions. Mol Cell Endocrinol 2010; 321:77-85.
- 142. Nikiforov YE, Yip L, Nikiforova MN. New strategies in diagnosing cancer in thyroid nodules: impact of molecular markers. Clin Cancer Res 2013; 19:2283-8.
- 143. Nikiforova MN, Wald AI, Roy S, Durso MB, Nikiforov YE. Targeted next-generation sequencing panel (Thyro Seq) for detection of mutations in thyroid cancer. J Clin Endocrinol Metab 2013; 98:E1852-60.
- 144. Kakudo K, Bai Y, Liu Z, Li Y, Ito Y, Ozaki T. Classification of thyroid follicular cell tumors: with special reference to borderline lesions. Endocr J 2012; 59:1-12.
- 145. Kakudo K, Mori I, Liu Z, Hui Z, Kakudo M. Papillary microcarcinoma and microtumor of the thyroid gland. J Basic Clinic Med 2013; 2:1-6.
- 146. Kakudo K, Wakasa T, Kakudo M, Liu Z: Borderline and precursor lesions of thyroid neoplasms. J Bas Clin Med 2015; 4:2-7.